

Mechanistic Studies of a Peptidic GRP78 Ligand for Cancer Cell-Specific Drug Delivery

Ying Liu,[†] Sebastian C. J. Steiniger,[†] YoungSoo Kim,[†] Gunnar F. Kaufmann,[†]
Brunhilde Felding-Habermann,[‡] and Kim D. Janda^{*,†,§}

*The Skaggs Institute for Chemical Biology and Departments of Chemistry and Immunology,
Worm Institute of Research and Medicine (WIRM), and Department of Molecular and
Experimental Medicine, The Scripps Research Institute, La Jolla, California 92037*

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Abstract: Major obstacles in the development of new therapeutic anticancer drugs are the low bioavailability of hydrophilic substances and the nonspecific toxicity toward healthy tissues. As such, cell-targeting oligopeptides have emerged as attractive drug delivery vehicles for a variety of different types of cargo. The recently identified peptide Pep42 binds to the glucose-regulated protein 78 (GRP78), which is overexpressed on the cell surface of human cancer cells and internalizes into these cells. Herein, we demonstrate how Pep42 can be utilized as a carrier for different types of cytotoxic drugs to specifically target human cancer cell lines *in vitro* in a strictly GRP78-dependent manner. Furthermore, the mechanism of internalization of Pep42 was elucidated and found to involve clathrin-mediated endocytosis. Pep42 subsequently colocalizes within the lysosomal compartment. Importantly, we also provide evidence that Pep42-conjugated quantum dots have the ability to selectively enrich in tumor tissue in a xenograft mouse model. Our results suggest that the highly specific GRP78–Pep42 interaction can be utilized for the generation of Pep42–drug conjugates as a powerful anticancer drug delivery system that could substantially enhance the efficacy of chemotherapy by increasing the drug–tumor specificity, thus minimizing the adverse side effects associated with conventional cancer therapeutics.

Keywords: Drug delivery; tumor targeting; GRP78; heat shock protein; cyclic peptide

Introduction

The lack of specificity seen with anticancer drugs in chemotherapy toward tumor tissues can result in adverse side effects, thereby limiting their clinical applications.¹ To overcome this obstacle, methods of delivering cytotoxic drugs specifically into the tumor tissue have to be investigated.² A promising therapeutic approach is the conjugation of a

cytotoxic drug to a ligand that recognizes a well-characterized cell surface marker, which is exclusively or at least preferentially expressed on tumor cells. Oligopeptides are candidates for such drug delivery purposes and have been employed successfully as targeting ligands.^{3–6} They can be chemically synthesized in large quantities, have low innate

* Address correspondence to this author. Mailing address: Department of Chemistry, The Scripps Research Institute, 10550 North Torrey Pines Road, La Jolla, CA 92037. Tel: 858 784-2516. Fax: 858 784-2590. E-mail: kdjanda@scripps.edu.

[†] The Skaggs Institute for Chemical Biology and Departments of Chemistry and Immunology.

[‡] Department of Molecular and Experimental Medicine.

[§] Worm Institute of Research and Medicine (WIRM).

(1) Langerak, A. D. *Chemotherapy Regimens and Cancer Care*; Landes Bioscience: Georgetown, TX, 2001; p 209.

(2) Zhou, X.; Chang, Y. C.; Oyama, T.; McGuire, M. J.; Brown, K. C. *J. Am. Chem. Soc.* **2004**, *126* (48), 15656–7.

(3) Torchilin, V. P.; Levchenko, T. S. *Curr. Protein Pept. Sci.* **2003**, *4* (2), 133–40.

(4) Lewin, M.; Carlesso, N.; Tung, C. H.; Tang, X. W.; Cory, D.; Scadden, D. T.; Weissleder, R. *Nat. Biotechnol.* **2000**, *18* (4), 410–4.

(5) El-Andaloussi, S.; Holm, T.; Langel, U. *Curr. Pharm. Des.* **2005**, *11* (28), 3597–611.

(6) Nahrendorf, M.; Jaffer, F. A.; Kelly, K. A.; Sosnovik, D. E.; Aikawa, E.; Libby, P.; Weissleder, R. *Circulation* **2006**, *114* (14), 1504–11.

toxicity, are readily amenable to derivatization, and are able to penetrate solid tumors.⁷ A number of studies have shown that peptides are able to deliver a wide range of bioactive molecules such as proteins, oligonucleotides, siRNAs, nanoparticles, and other large particles such as liposomes to a variety of cell lines both *in vitro* and *in vivo*.^{3–5,8,9}

The panning of phage display peptide libraries against cells, often referred to as biopanning, has proven successful for the isolation of peptidic ligands.^{10–12} However, these identified peptides typically did not possess any intrinsic cell type specificity, and only recently has the identification of cell type-specific peptides been reported.^{13,14} These peptides display certain characteristics of cell-penetrating peptides (CPPs) but, in contrast to all other previously described CPPs, demonstrate a restricted cell recognition pattern. Our laboratory has developed phage display biopanning protocols to identify peptides of constrained origin that internalize into tumor cells.¹⁵ Recently, we reported the selection and characterization of Pep42, a cyclic 13-mer oligopeptide,

CTVALPGGYVRVC, that specifically binds to glucose-regulated protein 78 (GRP78), internalizes into highly metastatic melanoma cells, and eventually remains inside the cell.¹⁶ In cancer cells, the overexpression of GRP78, an intracellular chaperone and member of the heat shock protein 70 (HSP70) family, provides a protective cellular response against stress conditions. However, this GRP78 overexpression represents a potentially attractive target as it results in the presence of GRP78 molecules on the cell surface of these cancer cells.^{17–19} Indeed, the presence of GRP78 on the cell surface has been demonstrated for a number of tumors, e.g.,

malignant human breast and lung cancers, thus making GRP78 a pan-neoplastic antigen.^{20–23}

Herein, we provide mechanistic investigations into a cancer cell-specific drug delivery approach exploiting tumor cell-specific expression of GRP78 using our recently identified GRP78 ligand, Pep42. In total, 3 cancer cell lines were evaluated for their surface expression level of GRP78. The specificity of the Pep42–GRP78 binding interactions were also examined using GRP78 siRNA experiments, which demonstrated that GRP78 is the singular cellular receptor for Pep42. In order to increase the efficacy of the Pep42–drug conjugates, the mechanism of internalization as well as the intracellular fate of Pep42 was investigated using live cell confocal fluorescence microscopy. These experiments established that Pep42 after binding to GRP78 is taken up via receptor-mediated endocytosis and ultimately locates into the lysosomal compartment of the cell. To evaluate the potential of Pep42 for the delivery of different cargo types into cancer cells, two cytotoxic Pep42–drug conjugates were generated, one based on the chemical fusion of Pep42 to the apoptosis-inducing oligopeptide D-(KLAKLAK)₂, the other consisting of a photosensitizer, hematoporphyrin, chemically conjugated to Pep42. Excitingly, both drug conjugates were able to exert their cytotoxic effects on a number of cancer cells in a GRP78-dependent manner, thus again underscoring the specificity of the Pep42–GRP78 interaction. Importantly, the ability of Pep42–nanoparticle conjugates to home into tumor tissue was demonstrated in an *in vivo* xenograft mouse model, which further confirmed the potential of GRP78 ligands as new cancer drug delivery vehicles.

Experimental Methods

Cell Lines and Culture Conditions. Human melanoma cells Me6652/4 and SJSA-1 osteosarcoma cells were cultured in RPMI 1640 medium (Invitrogen, Carlsbad, CA) supplemented with 10% fetal bovine serum (FBS, Invitrogen). HepG2 hepatoma cells and a normal human lung fibroblast cell line, CCD-11Lu (both from ATCC, Manassas, VA), were grown in Eagle's minimum essential medium (EMEM, Invitrogen) supplemented with 10% FBS, 1.0 mM sodium pyruvate, 2 mM L-glutamine, 1.5 g/L sodium bicarbonate, and 0.1 mM nonessential amino acids. A549 lung adenocarcinoma cells (ATCC) were maintained in Ham's F12K medium (ATCC) supplemented with 10% FBS, 2 mM

- (7) De, J.; Chang, Y. C.; Samli, K. N.; Schisler, J. C.; Newgard, C. B.; Johnston, S. A.; Brown, K. C. *Mol. Biosyst.* **2005**, *1* (2), 149–157.
- (8) Jarver, P.; Langel, U. *Drug Discovery Today* **2004**, *9* (9), 395–402.
- (9) Jarver, P.; Langel, U. *Biochim. Biophys. Acta* **2006**, *1758* (3), 260–3.
- (10) Moss, J. A.; Lillo, A.; Kim, Y. S.; Gao, C.; Ditzel, H.; Janda, K. D. *J. Am. Chem. Soc.* **2005**, *127* (2), 538–9.
- (11) Landon, L. A.; Deutscher, S. L. *J. Cell. Biochem.* **2003**, *90* (3), 509–17.
- (12) McGuire, M. J.; Samli, K. N.; Johnston, S. A.; Brown, K. C. *J. Mol. Biol.* **2004**, *342* (1), 171–82.
- (13) Vives, E. *J. Controlled Release* **2005**, *109* (1–3), 77–85.
- (14) Ruoslahti, E.; Duza, T.; Zhang, L. *Curr. Pharm. Des.* **2005**, *11* (28), 3655–60.
- (15) Gao, C.; Mao, S.; Ditzel, H. J.; Farnaes, L.; Wirsching, P.; Lerner, R. A.; Janda, K. D. *Bioorg. Med. Chem.* **2002**, *10* (12), 4057–65.
- (16) Kim, Y.; Lillo, A. M.; Steiniger, S. C.; Liu, Y.; Ballatore, C.; Anichini, A.; Mortarini, R.; Kaufmann, G. F.; Zhou, B.; Felding-Habermann, B.; Janda, K. D. *Biochemistry* **2006**, *45* (31), 9434–9444.
- (17) Arap, M. A.; Lahdenranta, J.; Mintz, P. J.; Hajitou, A.; Sarkis, A. S.; Arap, W.; Pasqualini, R. *Cancer Cell* **2004**, *6* (3), 275–84.
- (18) Kaufman, R. J. *Genes Dev.* **1999**, *13* (10), 1211–33.
- (19) Lee, A. S. *Trends Biochem. Sci.* **2001**, *26* (8), 504–10.

- (20) Misra, U. K.; Deedwania, R.; Pizzo, S. V. *J. Biol. Chem.* **2005**, *280* (28), 26278–86.
- (21) Fernandez, P. M.; Tabbara, S. O.; Jacobs, L. K.; Manning, F. C.; Tsangaris, T. N.; Schwartz, A. M.; Kennedy, K. A.; Patierno, S. R. *Breast Cancer Res. Treat.* **2000**, *59* (1), 15–26.
- (22) Bini, L.; Magi, B.; Marzocchi, B.; Arcuri, F.; Tripodi, S.; Cintorino, M.; Sanchez, J. C.; Frutiger, S.; Hughes, G.; Pallini, V.; Hochstrasser, D. F.; Tosi, P. *Electrophoresis* **1997**, *18* (15), 2832–41.
- (23) Uramoto, H.; Sugio, K.; Oyama, T.; Nakata, S.; Ono, K.; Yoshimastu, T.; Morita, M.; Yasumoto, K. *Lung Cancer* **2005**, *49* (1), 55–62.

L-glutamine, and 1.5 g/L sodium bicarbonate. Normal human dermal fibroblasts (HDFa) from adult skin (Cascade Biologics, Portland, OR) were cultured in Medium 106 supplemented with low serum growth supplement (LSGS). All cell cultures were maintained at 37 °C in a humidified atmosphere of 5% CO₂ in air.

Synthesis of Peptides and Peptide Conjugates. All peptides were prepared by stepwise solid-phase protocols for Fmoc chemistry using DIC/HOBt as previously described.^{16,24} Pep42–(Ac-HN-CTVALPGGYVRKC-CONH₂)–fluorescein isothiocyanate (Pep42–FITC) and Pep42–Bpa–biotin syntheses have been described; while Pep42–D-(KLAKLAK)₂ and D-(KLAKLAK)₂ were synthesized in a similar manner.¹⁶ A scrambled peptide with the sequence Ac-HN-CGVTPVGARYLK-CONH₂ and a linear Pep42 with the sequence Ac-HN-MTVALPGGYVRKM-CONH₂ were also synthesized. Unrelated sequences (H₂N-ACDEYIRVWYARHC AhxK (FITC)-CONH₂ or H₂N-ACGWDGVDYMSDWVC AhxK (FITC) -CONH₂) from the same pIX cyclic peptide library were used as negative controls. Pep42–dPEG6–hematoporphyrin was prepared by conjugating hematoporphyrin (Sigma, St. Louis, MO) with PEG linkers (Fmoc-amido-dPEG6-acid from Quanta BioDesign, Ltd., Powell, OH) on the free lysine side chain of Pep42 using 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDCI) and DMSO for 24 h at room temperature (RT). RP-HPLC was performed employing binary gradients of solvents A and B, where A is 0.1% TFA in water and B is 0.09% TFA in acetonitrile. Analytical RP-HPLC was performed using a Vydac 214TP5415 column at a flow rate of 1 mL/min, with detection at 214 nm during a linear gradient of 20–80% B over 30 min. Preparative RP-HPLC was performed using a Vydac 214TP101522 column at a flow rate of 10 mL/min, with detection at 220 nm during a linear gradient of 25–55% B over 30 min. In all cases, fractions were analyzed off-line using an ABI/Sciex 150EX single quadrupole mass spectrometer and judged for purity after a consistent summing of 20 scans in multichannel analysis (MCA) mode, using the [M + 2]²⁺ charged species. For preparative purification purposes, fractions that contained no single [M + 2]²⁺ charged species which accounted for more than 10% of the total ion intensity were designated “pure” and pooled; the homogeneity of this pool was verified by analytical RP-HPLC and was >96%. Concentrations of peptides or peptide conjugates were determined using a UV–vis absorption spectrometer. Pep42–FITC: 490 nm (ϵ = 68 000). Pep42 or Pep42–Bpa–biotin: 280 nm (ϵ = 1280). Pep42–D-(KLAKLAK)₂: 280 nm (ϵ = 1400). Pep42–dPEG6–hematoporphyrin or hematoporphyrin: 390 nm (ϵ = 79 400).

Cell-Free GRP78 Binding Assay. GRP78 (Stressgen, San Diego, CA), HSP70 (Stressgen), and BSA (Bio-Rad, Richmond, CA) (10 μ g/mL, 25 μ L/well) were immobilized on 96-well microtiter wells overnight at 4 °C. After blocking with 1% skim milk, 10⁹ colony forming units (cfu) of

peptide-phage (Pep42-phage or an unrelated peptide-phage as negative control) in 25 μ L of PBS/1% skim milk was added into each well for 1 h at RT with slow agitation. For detection of peptide-phage, wells were washed with PBS and bound phage clones were recovered by infection with host bacteria in log phase ER2537, New England Biolabs, Ipswich, MA). Aliquots of the bacterial culture were seeded on a SOBAG plate and incubated overnight at 37 °C.¹⁶ The phage cfu was determined in triplicate. For Pep42 binding assays, increasing concentrations of Pep42 or a scrambled Pep42 were added to wells coated with GRP78 and incubated for 30 min at 37 °C, followed by an incubation step with 10⁹ cfu of Pep42-phage for 1 h at RT.

Identification of GRP78 on the Cell Surface of Human Cancer Cells. Expression of surface GRP78 on cells was assessed by flow cytometry. Cells were dissociated with Versene 1:5000 (Invitrogen) and resuspended in Opti-MEM I reduced-serum medium (Invitrogen). Approximately 1 \times 10⁶ cells were harvested, washed, and incubated with a saturating amount (10 μ g/mL) of mouse anti-GRP78 or a monoclonal anti- β -actin antibody (Sigma) as a negative control in staining buffer (PBS pH 7.4, 4% FBS and 0.1% NaN₃) for 1 h at 4 °C. Cells were washed and stained with FITC-conjugated anti-mouse IgG secondary antibody for 45 min at 4 °C. Additional washing (2 \times) with cold FCM washing buffer (Santa Cruz Biotechnology, Santa Cruz, CA) was followed by propidium iodide (Invitrogen) as a counterstain. Fluorescence intensity of the samples was measured on a FACSsort (BD Biosciences, San Diego, CA) using Cellquest and FlowJo software, and at least 5 \times 10³ cells were analyzed for each sample.

Cell Surface Binding Analysis Using Flow Cytometry. Cultured cells were dissociated, washed in culture medium, and aliquoted (\sim 1 \times 10⁵–10⁶ cells/tube) into culture tubes. Opti-MEM I reduced serum medium was renewed 1 h at 4 °C prior to the addition of the peptide. To the cells was added either 50 nM FITC-conjugated Pep42 or a FITC-conjugated unrelated peptide as negative control followed by incubation for 25 min at 4 °C. The cells were washed (2 \times) with a cold FCM washing buffer; this was followed by a propidium iodide counterstain and FACS analysis. A goat polyclonal anti-GRP78 antibody (Santa Cruz Biotechnology) was used to evaluate the competitive inhibition of Pep42–FITC or control peptide–FITC binding.

Western Blot Analysis. Cells were lysed in RIPA buffer (150 mM NaCl, 10 mM Tris-HCl, pH 7.4, 5 mM EDTA, 1% Triton X-100), supplemented with 1:100 protease inhibitor cocktail (Sigma) and 1:100 phosphatase inhibitor cocktail set II (Calbiochem, San Diego, CA). Cell lysates were centrifuged at 10000g for 20 min. The protein concentration in the supernatant was determined by Bradford assay (Bio-Rad). Aliquots containing 20–50 μ g of protein were resolved on sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS–PAGE), followed by electrotransfer to a nitrocellulose membrane. Immunodetection was carried out using Streptavidin-horseradish peroxidase (HRP) (Amersham Biosciences,

(24) Chan, W. C.; White, P. D. *Fmoc Solid Phase Peptide Synthesis*; Oxford University Press: 2000.

Piscataway, NJ), Hsp70/Hsc70 antibody (Stressgen), or Bip/GRP78 antibody (BD Biosciences).

Transfection of Cells with Small Interfering RNA (siRNA) Oligomers. The siRNA used against human GRP78 (Santa Cruz Biotechnology) was a pool of 3 target-specific 20–25 nt siRNAs that were designed to knock down gene expression. Cells were thus transfected with GRP78 siRNA or control siRNA (Santa Cruz) using Lipofectamine 2000 reagent (Invitrogen), and the transfection protocol was provided with the kit. After 48 h, Western blot analysis and cell viability assays were carried out.

Fluorescence Microscopy. Quantum dots (QDots 525, Quantum Dot Corp., Hayward, CA) were coupled to Pep42 as previously detailed.¹⁶ For treatment with fluorescent proteins, A549 cells were grown on a 35 mm glass bottom dish to approximately 70% confluence. After a 30 min incubation period at 37 °C in serum-free Ham's F12K medium, 6 μ M Pep42–quantum dots (QDots) were added 1 h before the addition of the fluorescent proteins. Final concentrations of fluorescent materials were 10 μ g/mL Alexa Fluor (AF)-594-conjugated transferrin, 10 μ g/mL AF-594 labeled cholera toxin subunit B (recombinant), 25 nM Mitotracker Red, 20 nM ERtracker Red, and 12 nM LysoTracker Red. All reagents except the QDots were obtained from Invitrogen. At different time points as indicated in each figure, media were changed with PBS and colocalization experiments in live cells were imaged using a laser scanning confocal microscope (MRC1024, Bio-Rad, Hercules, CA). Weighted colocalized coefficients were estimated using Zeiss LSM Image examiner software. For inhibitor-treated experiments, chlorpromazine, nystatin, and methyl- β -cyclodextrin (MCD) were purchased from Sigma-Aldrich. A549 cells were pretreated with 25 μ M nystatin, 30 μ M chlorpromazine, or 5 mM methyl- β -cyclodextrin (MCD) for 10 min, and then Pep42–QDots, AF-594–transferrin, or AF-594–cholera toxin subunit B was added, respectively.

Cell Viability Assay. Cell suspensions of 1×10^4 cells per well were seeded in 96-well plates for 18 h in medium containing 10% FBS. Before the assay was carried out, culture conditions were changed to low-serum medium (1% FBS), after which the cells were incubated with increasing concentrations of the selected peptides including Pep42–D-(KLAKLAK)₂ or (Pep42 and free D-(KLAKLAK)₂) in 100 μ L for the indicated time at 37 °C. Cell viability was measured with a cell proliferation reagent (Promega, St. Luis Obispo, CA). For the evaluation of the porphyrin phototoxicity, cells were prepared as described (*vide supra*) and treated with varying concentrations of Pep42–hematoporphyrin or free hematoporphyrins at 37 °C, respectively. After the indicated time as designated in each figure, the medium was replaced with 50 mM HEPES (pH 7.4) and the cells were exposed to visible light for 1 h at RT 24. Upon completing of photodynamic treatment the cells were incubated for 24 h at 37 °C, and the phototoxicity was detected as described (*vide supra*).

Apoptosis Assay Using Flow Cytometry. Apoptotic cells were stained with Annexin V–FITC and propidium iodide (Invitrogen) according to the manufacturer's instructions and analyzed by flow cytometry. Cell debris was excluded from analysis by appropriate forward light scatter threshold settings. Cells that stained with Annexin V–FITC, but not propidium iodide, were regarded as early apoptotic cells.²⁵

Establishment of Tumor-Bearing Mice. The ability of peptide-coated quantum dots to home to their targets *in vivo* was tested in xenograft mice. To establish tumor xenografts, 10⁶ exponentially growing melanoma Me6652/4 tumor cells were injected intradermally in the flank of CB17Scid male mice (Taconic Farms, Germantown, NY). The mice were used for *in vivo* targeting experiments 4 weeks after the tumor cell inoculation. Peptide-coated quantum dots preparations (100–200 μ g in 0.1–0.2 mL of PBS) were injected into the lateral tail vein of a mouse and allowed to circulate for 120 min. While still under anesthesia, the mouse was perfused with 4% paraformaldehyde through the heart. The organs were harvested and frozen in Tissue Tek OCT embedding medium (Sakura Finetek, Torrance, CA) before sectioning. The sections were mounted with Vectashield mounting medium with or without DAPI (Vector Laboratories, Burlingame, CA) to visualize cell nuclei before examination under an inverted fluorescent or confocal microscope. The anti-CD31 antibody (10 μ g/mL; BD Biosciences) applied on the tissue sections was detected with Alexa Fluor 594-conjugated secondary antibody.

Results

GRP78 Is the Exclusive Cell Surface Receptor for Pep42.

Applying phage display technology our laboratory has identified the cyclic peptide Pep42, CTVALPG-GYVRVC, that specifically recognizes GRP78 present on the cell surface of melanoma Me6652/4 cells, and is subsequently internalized.¹⁶ For tumor site-specific therapy it is of paramount importance that the targeted molecular feature is indeed unique, or primarily restricted to cancer cells and absent on normal cells. It had been previously shown that a number of human primary and cancer cell lines overexpress GRP78, resulting in the cell membrane localization of this protein; thus, we reasoned that GRP78 might represent an attractive target for tumor cell-specific cytotoxic therapy.^{17,26}

In order to study the mechanism of internalization and the *in vivo* localization of Pep42, we have synthesized the peptide by replacing the valine at position 12 with a lysine residue to facilitate the coupling of conjugates.¹⁶ We note that since no functional differences between this lysine-bearing derivative and the original Pep42 sequence were

(25) Katsuta, M.; Takigawa, Y.; Kimishima, M.; Inaoka, M.; Takahashi, R.; Shiohara, T. *J. Immunol.* **2006**, 176 (12), 7736–44.

(26) Shin, B. K.; Wang, H.; Yim, A. M.; Le, Naour, F.; Brichory, F.; Jang, J. H.; Zhao, R.; Puravs, E.; Tra, J.; Michael, C. W.; Misk, D. E.; Hanash, S. M. *J. Biol. Chem.* **2003**, 278 (9), 7607–16.

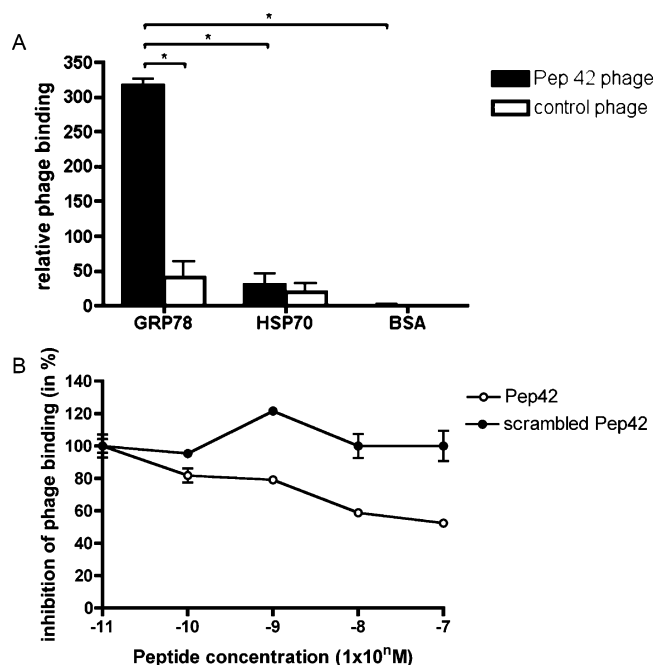


Figure 1. Pep42-displaying phage particles bind specifically to GRP78. (A) GRP78, HSP70, and BSA were coated on microtiter wells at 10 μ g/mL and incubated with either Pep42-phage or control phage. The control phage displays an unrelated sequence. Phage input was 10^9 cfu per well. Results are expressed as mean \pm standard error of the mean (SEM) of triplicate wells. (B) The inhibition of binding of Pep42-phage to immobilized GRP78 was obtained by the addition of increasing concentrations of Pep42 or a scrambled Pep42. *Student's *t* test, $p < 0.05$.

observed, we have termed this sequence “Pep42” for the sake of simplicity (Supporting Information, Figure S1).

To unequivocally probe the exclusivity of the binding interactions between Pep42 and GRP78, we evaluated its ability to discriminate between recombinant GRP78 protein and HSP70, another member of the HSP family that shares a high degree of protein sequence homology with GRP78, as well as to bovine serum albumin (BSA) as a surrogate for serum proteins in a cell-free system. The Pep42-phage indeed bound specifically only to immobilized GRP78 while no binding was observed to either control protein (Figure 1A). Furthermore, the binding of Pep42 to GRP78 was inhibited in a dose-dependent manner by the addition of free Pep42, while the addition of scrambled control peptide did not affect the binding of phage to GRP78 (Figure 1B). These results demonstrated that Pep42 specifically recognizes and binds to GRP78.

The binding of Pep42 to GRP78 overexpressed on the surface of cancer cells was also analyzed. The presence of GRP78 on the cancer cell surface has only recently been appreciated, while GRP78 is typically absent on the cell membrane of normal cells.²⁰ Indeed, a global profiling approach of the cell surface proteome of cancer cells uncovered that GRP78 is a cell membrane protein on a number of cancer cells, thus making GRP78 a pan-neoplastic antigen.²⁶ To evaluate Pep42's ability to bind to human

cancer cell lines other than Me6652/4, we analyzed the expression of surface GRP78 in a range of cancer cell lines including lung adenocarcinoma cells (A549), osteosarcoma cells (SJSA-1), hepatoma cells (HepG2), and two normal control fibroblast cell lines via flow cytometry using a monoclonal antibody (mAb) directed against human GRP78. A shift of fluorescence was observed for HepG2, SJSA-1, and A549 cells, while no shift could be seen using the normal skin fibroblast cell line, HDFa, and a normal lung fibroblast cell line, CCD-11Lu (Supporting Information, Figure S2), indicating the low expression of GRP78 on the surface of these two control cell lines.

The ability of Pep42 to selectively bind to the positively identified GRP78-expressing cell lines was investigated with Me6652/4, HepG2, A549, and SJSA-1 cells as well as control HDFa cells. The cells were incubated with 50 nM FITC-labeled Pep42 or a FITC-labeled control peptide for 25 min at 4 $^{\circ}$ C, and cell fluorescence was measured by flow cytometry. As shown in Figure 2A, Pep42 bound to GRP78 on the surface of Me6652/4, HepG2, A549, and SJSA-1 cells, while only minimal recognition was observed for HDFa cells. Based on these GRP78 expression data and Pep42 binding results, the cancer cell lines were used interchangeably.

A photoaffinity labeling experiment generally used to analyze ligand–receptor interactions was also conducted in analogy to our previous report that led to the identification of the Pep42 receptor on Me6652/4 cells but now using HepG2 cells and HDFa cells.¹⁶ The data clearly demonstrated that Pep42 interacts with the same protein on HepG2 cells as it does on Me6652/4, but does not bind to any other surface protein on the control HDFa cells (Figure 2B).

An anti-GRP78 polyclonal antibody was utilized to block Pep42 on the cell surface of HepG2 cells, which were chosen due to their high level of GRP78 surface expression, and the binding of Pep42 was subsequently examined. The pretreatment of cells with the pAb significantly decreased the binding of Pep42 to HepG2 cells (Figure 2C). Last, GRP78 expression in Me6652/4 cells was suppressed by transfection with GRP78-specific siRNA. As shown in Figure 2D, the endogenous GRP78 protein level was significantly reduced in cells transfected with GRP78 siRNA but not in those treated with control siRNA. The GRP78-specific siRNA had no effect on the expression levels of control proteins. Importantly, Pep42 binding was notably reduced in cells treated with the GRP78-specific siRNA. In total, these results underscore that the binding of Pep42 to cells is strongly dependent on the cell surface expression level of GRP78, thus granting cancer cell-targeting specificity for Pep42.

Selective Induction of Apoptosis in GRP78-Overexpressing Tumor Cells Mediated by Pep42–D-(KLAK-LAK)₂. To evaluate the drug delivery capabilities of Pep42,

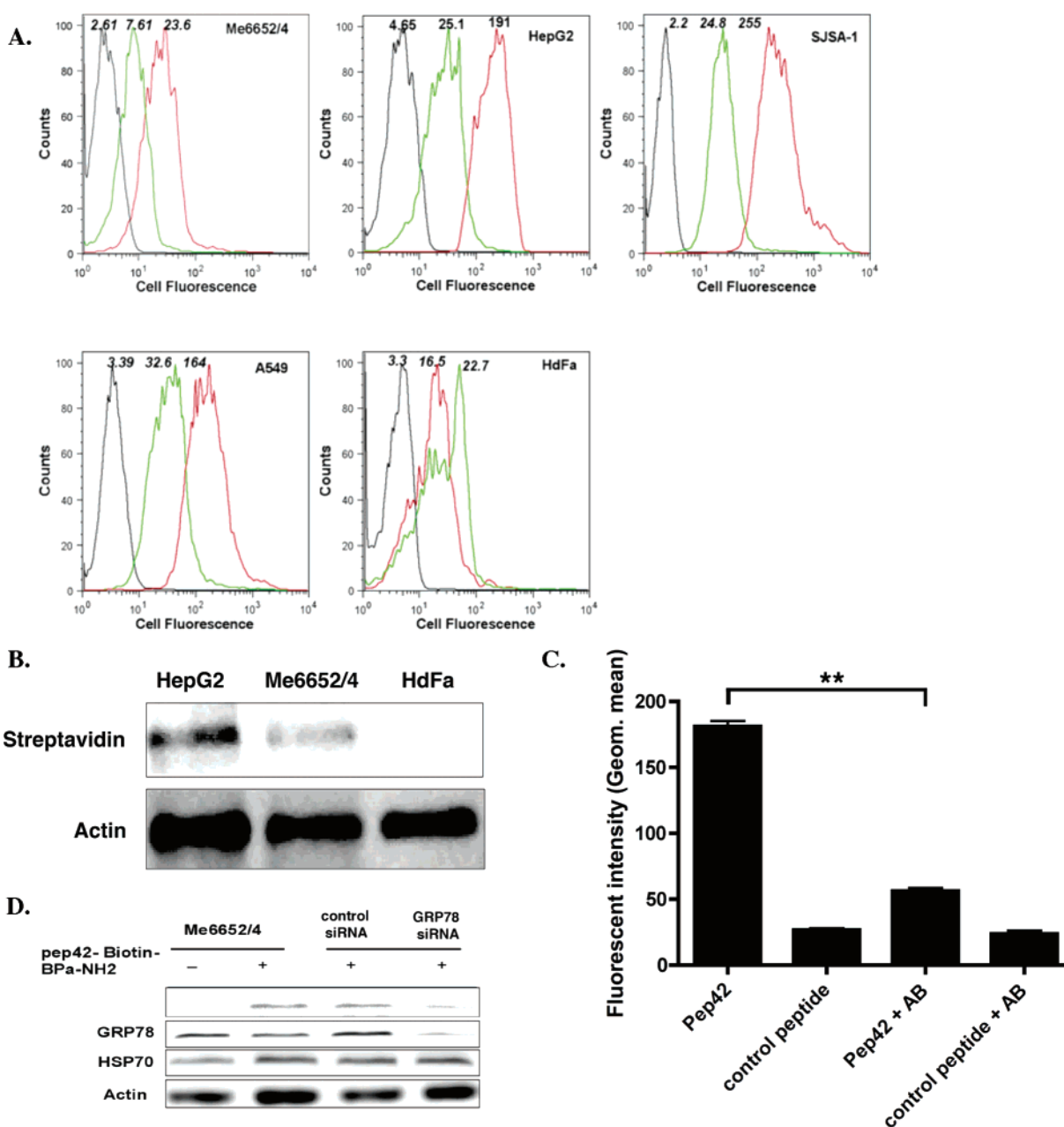


Figure 2. Pep42 binding to tumor cells is strictly dependent on GRP78 cell surface expression. (A) Cells were incubated with either 50 nM FITC-conjugated Pep42 (red curve) or FITC-conjugated control peptide (green curve) for 25 min at 4 °C with slow rotation. The black curves correspond to cells incubated in the absence of any peptide. (B) Pep42 binding to tumor cells was estimated using photoaffinity labeling. Western blot of Pep42 with its interacting proteins is shown. Here, cells were incubated with 50 μ M Pep42–Bpa–biotin for 1 h at 4 °C in the dark. The Pep42 protein interactions found on the cell surface were purified by avidin–agarose affinity chromatography, separated by SDS–PAGE, and transferred onto a nitrocellulose membrane. Upon incubation with streptavidin–HRP the membrane was developed with luminal and analyzed by chemoluminescence. (C) Inhibition of binding of Pep42 to HepG2 cells was evaluated by the addition of a monoclonal anti-GRP78 antibody and the subsequent incubation of cells with Pep42 or a control peptide. **Difference of cells treated with Pep42 only, Student's *t* test, $p < 0.01$. (D) Effect of silencing the GRP78 gene expression on binding of Pep42 to Me6652/4 cells. After 48 h of transfection with GRP78 siRNA or control siRNA, cells were incubated with Pep42–Bpa–biotin. The detection was carried out as described for Figure 2B.

in vitro cell killing assays were conducted utilizing conjugates of Pep42 chemically fused to the apoptosis-inducing peptide D-(KLAKLAK)₂.^{27,28} As previously noted the amphipathic peptide D-(KLAKLAK)₂ does not interact with the zwitterionic plasma membrane of eukaryotic cells, however,

it can disrupt the negatively charged mitochondrial membrane and, thus, subsequently induces apoptosis after internalization.²⁷

Pep42–D-(KLAKLAK)₂-mediated cytotoxicity was observed in the two GRP78-positive tumor cell lines analyzed,

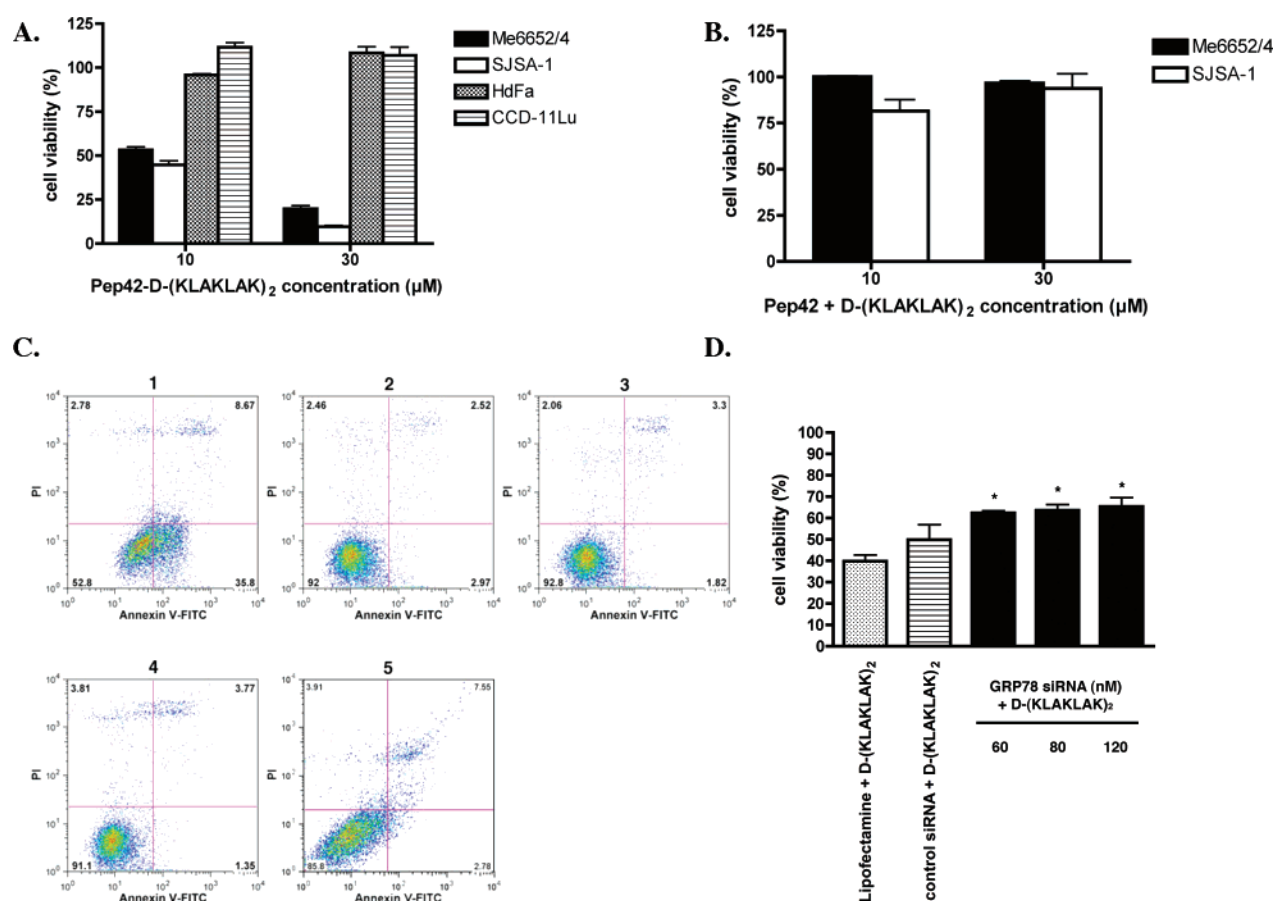


Figure 3. Selective killing and induction of apoptosis in surface GRP78 expressing cancer cells mediated by Pep42–D-(KLAKLAK)₂. (A) Cells were incubated with increasing concentrations of the peptides Pep42–D-(KLAKLAK)₂ or (B) a mixture of free Pep42 and D-(KLAKLAK)₂ in 100 μ L 1% FBS or supplemented medium for 24 h at 37 $^{\circ}$ C. The viability of the cells was assessed by a MTT assay. Results are expressed as percentages of cells still viable relative to nontreated cells. (C) SJSA-1 cells cultured in the presence of Pep42–D-(KLAKLAK)₂ or negative controls (free D-(KLAKLAK)₂, free Pep42, or scrambled Pep42) for 6 h at 37 $^{\circ}$ C were washed in cold PBS and resuspended in staining buffer prior to the addition of FITC-labeled Annexin V and propidium iodide for 15 min at room temperature. Flow cytometry was performed on SJSA-1 cells gated on the basis of their forward and side light scatter with any cell debris excluded from the analysis. Cell staining with Annexin V-FITC but no propidium iodide were regarded as early apoptotic cells. Early apoptotic cells (1) Pep42–D-(KLAKLAK)₂ 35.8%; (2) free Pep42 2.97%; (3) free D-(KLAKLAK)₂ 1.82%; (4) medium 1.35%; (5) scrambled Pep42–D-(KLAKLAK)₂ 2.78%. (D) Surface GRP78 expression was suppressed by GRP78 specific siRNA using Lipofectamine. After 48 h, cells were treated with 30 μ M Pep42–D-(KLAKLAK)₂ for 3 h at 37 $^{\circ}$ C. *Difference from Pep42–D-(KLAKLAK)₂ treated cells, Student's *t* test, *p* < 0.05.

Me6652/4 and SJSA-1 (Figure 3A).²⁹ In order to reduce nonspecific background and increase sensitivity, the assays were carried out in low serum containing media. Importantly, no cytotoxic effects were seen when these two cell lines were incubated with an equimolar mixture of Pep42 and free D-(KLAKLAK)₂ (Figure 3B). The GRP78-negative cell line was completely resistant to the peptide conjugate-mediated

cytotoxicity. Interestingly, the kinetics of the induction of apoptosis were slightly different depending on the cell line, suggesting a varying degree of susceptibility to toxicity in these cells (Supporting Information, Figure S3).

The evaluation of the extent of the apoptotic cell death induced by Pep42–D-(KLAKLAK)₂ in SJSA-1 cells was undertaken. Flow cytometric analysis using propidium iodide (PI) and Annexin V–FITC revealed that Pep42–D-(KLAKLAK)₂ induced an increase of Annexin V-positive cells, which represent cells in the early stages of apoptosis (Figure 3C). Moreover, we investigated the importance of surface GRP78 in the cell killing by Pep42–D-(KLAKLAK)₂. Indeed, the knockdown of GRP78 protein using siRNA methodology resulted in a significant reduction in the extent of cell death induced by Pep42–D-(KLAKLAK)₂, providing further evidence that surface GRP78 is required for the

(27) Marks, A. J.; Cooper, M. S.; Anderson, R. J.; Orchard, K. H.; Hale, G.; North, J. M.; Ganeshaguru, K.; Steele, A. J.; Mehta, A. B.; Lowdell, M. W.; Wickremasinghe, R. G. *Cancer Res.* **2005**, *65* (6), 2373–7.

(28) Mintz, P. J.; Kim, J.; Do, K. A.; Wang, X.; Zinner, R. G.; Cristofanilli, M.; Arap, M. A.; Hong, W. K.; Troncoso, P.; Logothetis, C. J.; Pasqualini, R.; Arap, W. *Nat. Biotechnol.* **2003**, *21* (1), 57–63.

(29) Elmquist, A.; Langel, U. *Biol. Chem.* **2003**, *384* (3), 387–93.

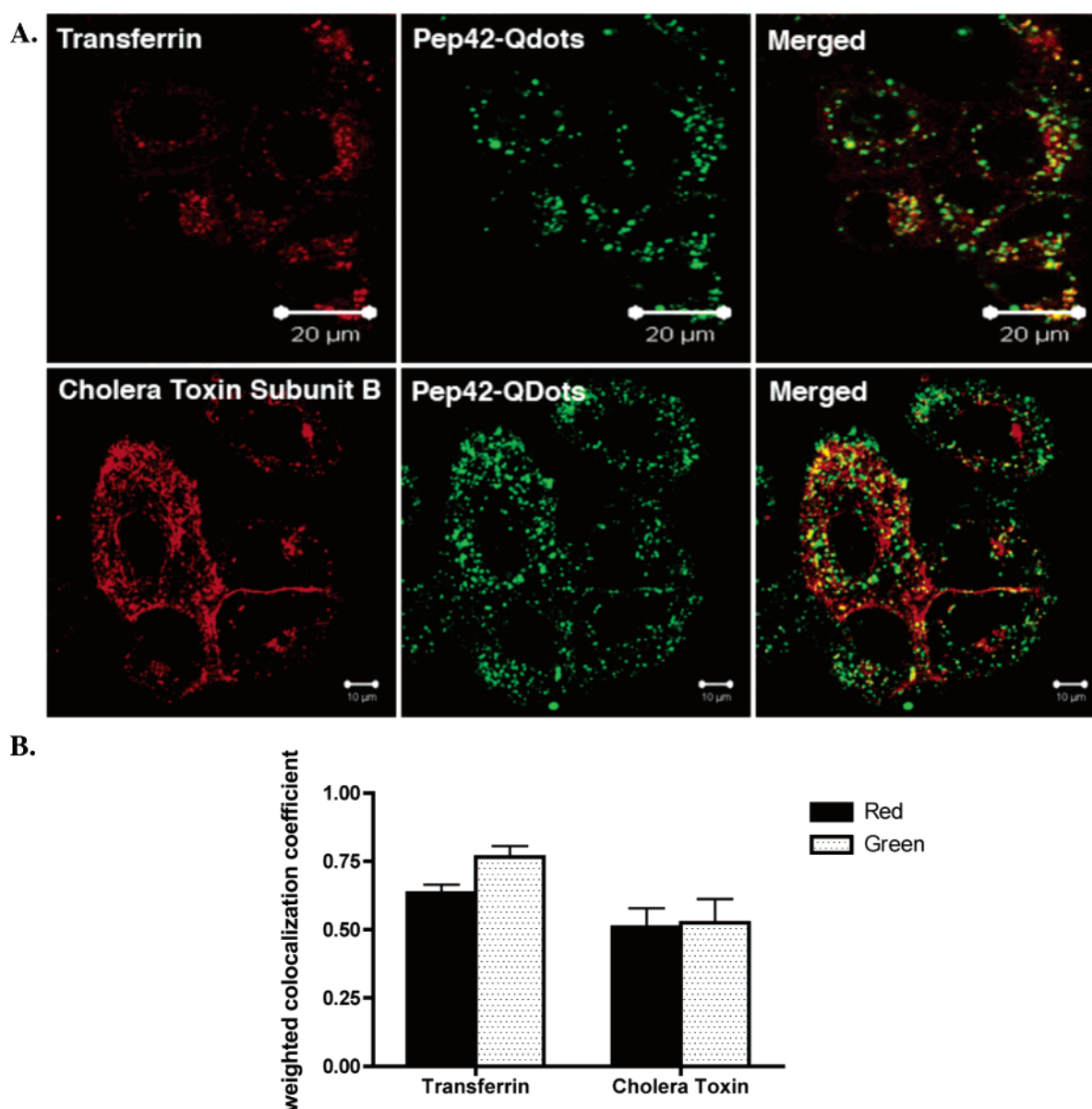


Figure 4. Pep42 partially colocalizes with known markers of endocytosis. (A) A549 cells were preincubated with Pep42–QDots 525 for 1 h at 37 °C, and then either 10 μ g/mL AF-594–transferrin or 10 μ g/mL AF-594–cholera toxin subunit B was added for 30 min at 37 °C. The cells were then washed with PBS and analyzed by laser scanning confocal microscopy. (B) Weighted colocalization coefficients were measured with Zeiss LSM Image examiner software. “Red” in the legend means the colocalized coefficient that red channel colocalizes with the green channel. “Green” in the legend means the colocalized coefficient that the green channel colocalizes with the red channel.

delivery of D-(KLAKLAK)₂ by Pep42 (Figure 3D). This siRNA experiment also indicated that the cell membrane presence of GRP78 on human cancer cells is solely dependent on the overexpression of the gene in cancer cells. Taken together the data strongly suggest that Pep42 could be an effective tumor cell-targeting drug delivery system.

Pep42/GRP78 Complex Is Internalized by Clathrin-Mediated Endocytosis. To take advantage of the full potential of Pep42 as a novel carrier for cancer cell targeting, it is important to elucidate the underlying mechanism that enables the cell-targeting peptide to internalize into cells and localize into a specific intracellular compartment. Endocytosis can occur through a number of mechanisms including

clathrin-mediated, caveolae/lipid raft-mediated, and clathrin- and caveolae-independent pathways.⁵ Transferrin is known to be internalized via clathrin-coated invaginations on the plasma membrane that detach to form clathrin-coated vesicles.³⁰ On the other hand, bacterial toxins, such as cholera toxin, utilize caveolae-mediated endocytosis to enter cells.³⁰

To assess the intracellular fate of Pep42 endosomes, we added either Alexa Fluor (AF)-594-labeled transferrin and Pep42–quantum dots (QDots) or AF-594-labeled cholera

(30) Fittipaldi, A.; Ferrari, A.; Zoppe, M.; Arcangeli, C.; Pellegrini, V.; Beltram, F.; Giacca, M. *J. Biol. Chem.* **2003**, 278 (36), 34141–9.

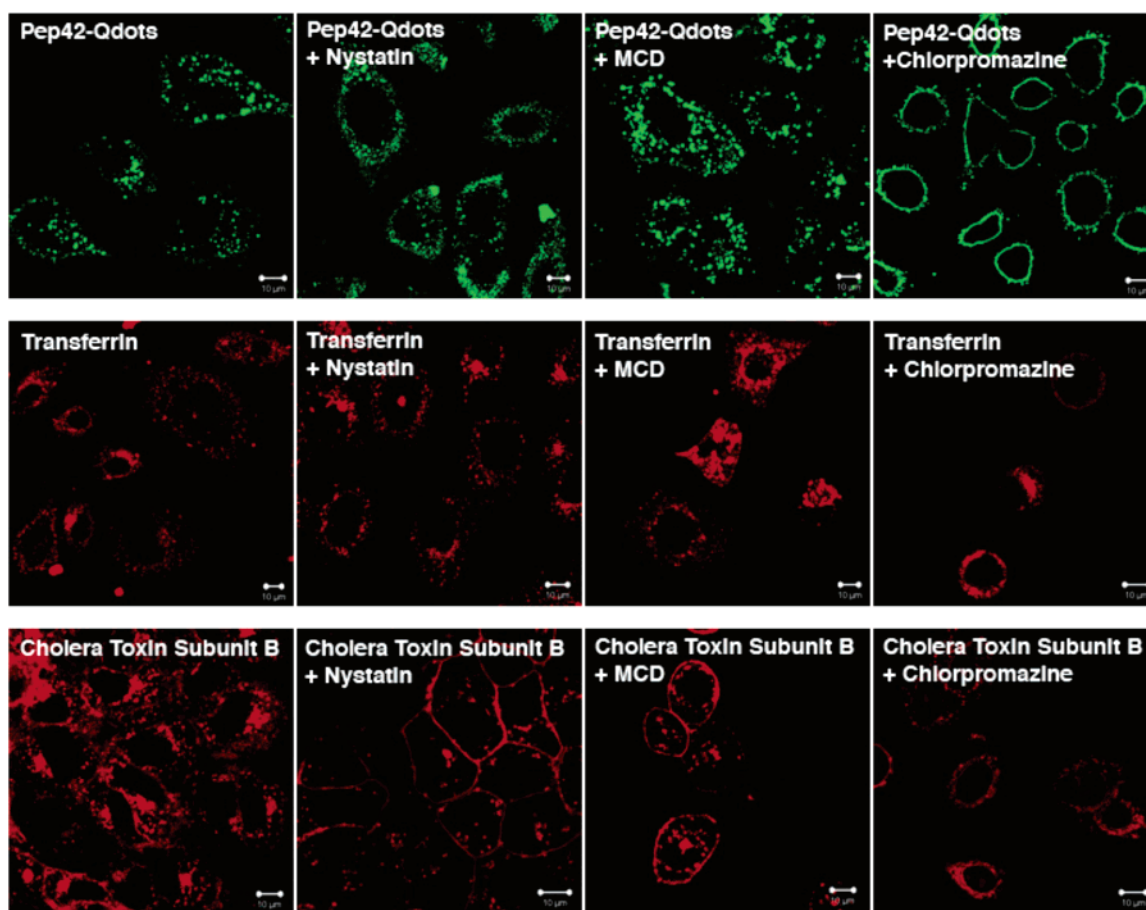


Figure 5. Inhibition of Pep42 uptake using known endocytosis inhibitors. A549 cells were pretreated with 25 μ M nystatin, 30 μ M chlorpromazine, or 5 mM methyl- β -cyclodextrin (MCD) for 10 min, and then Pep42–QDots 525, AF-594–transferrin, or AF-594–cholera toxin subunit B was added, respectively.

toxin subunit B (CTB) and Pep42–QDots to cultured A549 cells. As seen in Figure 4A, Pep42–QDots colocalized with transferrin and only to a lesser extent with CTB. The colocalization coefficients are also shown (Figure 4B).

To further elucidate the uptake mechanism, we next studied the effects of several inhibitors of endocytosis on the cellular uptake of Pep42 by A549 cells. Chlorpromazine, a known inhibitor of clathrin-mediated endocytosis, had the expected inhibitory effect on the uptake of Pep42 or transferrin, but no difference was observed for CTB. In contrast, the uptake of CTB was inhibited by the pretreatment of cells with nystatin or methyl- β -cyclodextrin (MCD), both inhibitors of caveolae-mediated endocytosis. As anticipated, neither nystatin nor MCD had any effect on the uptake of Pep42 or transferrin. There was a minor effect on cholera toxin internalization by chlorpromazine; however, this effect was very weak (Figure 5). The above results strongly indicate that the Pep42 uptake into A549 cells is clathrin-mediated but not caveolae-mediated endocytosis.

Endocytosis of clathrin-coated vesicles is also known to involve progressive endosome maturation and eventual fusion with the cell lysosome.³⁰ We assessed the colocalization of internalized Pep42–QDots with the organelle-specific fluorescent markers LysoTracker Red (lysosomes), Mitotracker

Red (mitochondria), and ERTracker Red (endoplasmic reticulum) (Figure 6A). Thus, 88% Pep42–QDots colocalized with lysosomes after 1 h incubation of Pep42–QDots. At the same time, approximately 73% lysosomes were targeted by internalized Pep42–QDots (Figure 6B) demonstrating that Pep42 preferentially localizes into the lysosomal compartment.

Pep42-Mediated Subcellular Transport of Porphyrin Conjugates. The biological efficacy of porphyrin-based sensitizers for photodynamic therapy of tumors relies mainly on their efficient translocation across cellular membranes and their delivery into specific organelles of cancer cells.³¹ It has been demonstrated that functionalized porphyrin conjugates bearing a fusogenic peptide localize mainly into lysosomes.³¹ Based on the colocalization data, it was important to prove that Pep42 can specifically target and effectively carry a drug into lysosomes of tumor cells; thus, a carboxylic acid functionalized hematoporphyrin was designed and chemically conjugated to Pep42. The phototoxicity of the Pep42–hematoporphyrin conjugate was evaluated using HepG2 and A549, cell lines that were not susceptible to Pep42–D-

(31) Sibrian-Vazquez, M.; Jensen, T. J.; Hammer, R. P.; Vicente, M. *G. J. Med. Chem.* **2006**, 49 (4), 1364–72.

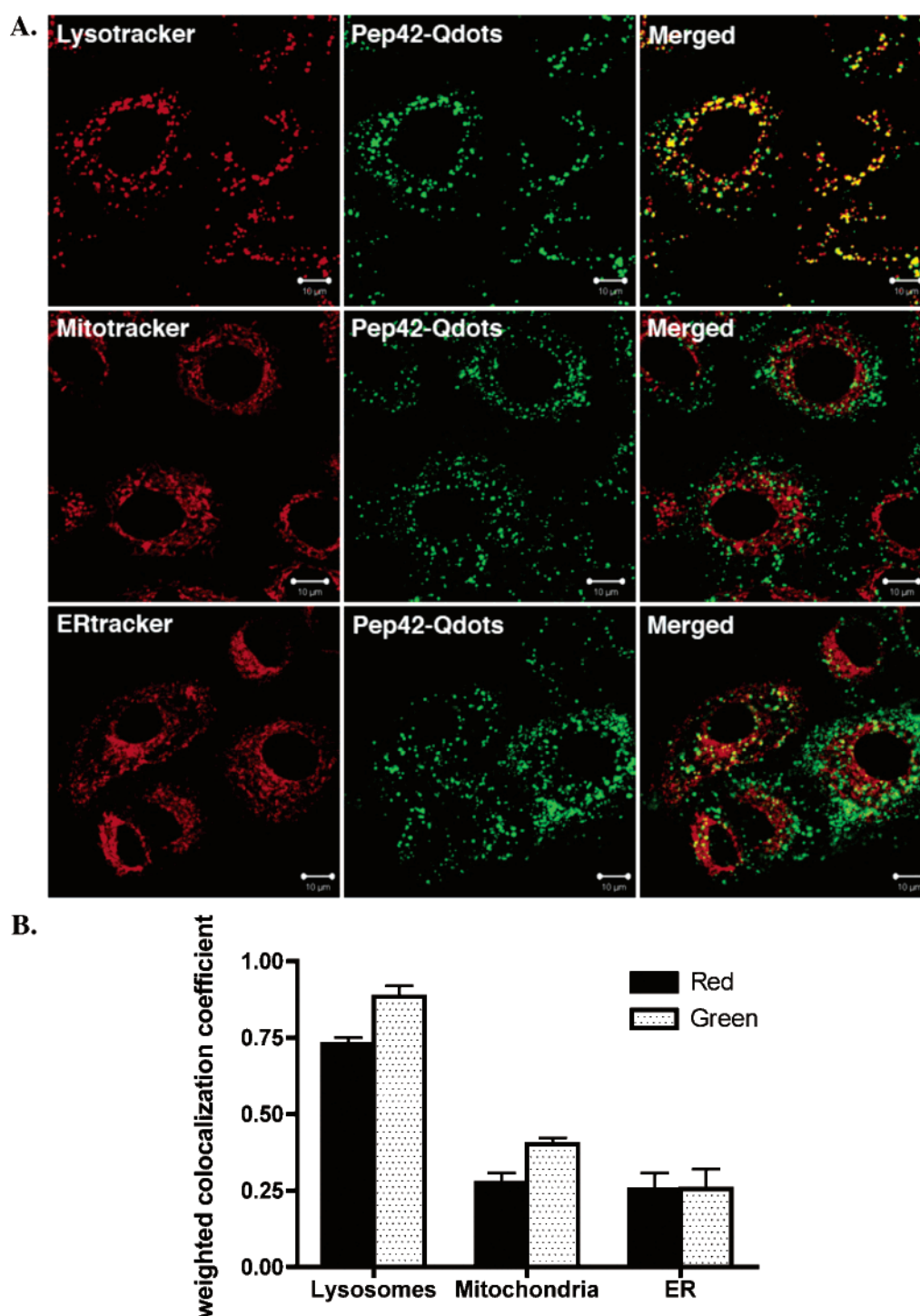


Figure 6. Pep42–QDot conjugates colocalize with the lysosomal compartment. (A) Colocalization of Pep42–QDots with Lysotracker Red in A549 cells. Cells were treated with 12 nM Lysotracker Red for 30 min, 20 nM ERtracker Red for 20 min, or 25 nM Mitotracker Red for 15 min after incubation of the peptide for 1 h at 37 °C. (B) Weighted colocalization coefficients were measured with Zeiss LSM Image examiner software. “Red” in the legend means the colocalized coefficient that red channel colocalizes with the green channel. “Green” in the legend means the colocalized coefficient that the green channel colocalizes with the red channel.

(KLAKLAK)₂ conjugate (Supporting Information, Figure S4). Cells were exposed to increasing concentrations (0–20 μ M) of Pep42-hematoporphyrin conjugate for 18 h, as shown in Figure 7A. The conjugate showed no toxicity at concentrations up to 10 μ M in the dark on A549 cells (data not

shown). However, the conjugate showed phototoxicity upon exposure to visible light for 1 h on HepG2 and A549 cells, while the surface GRP78-negative cell lines, HDFa and CCD-11Lu, were not affected by the conjugate. The IC₅₀ values for the conjugate were 9 μ M on HepG2 and 8 μ M on

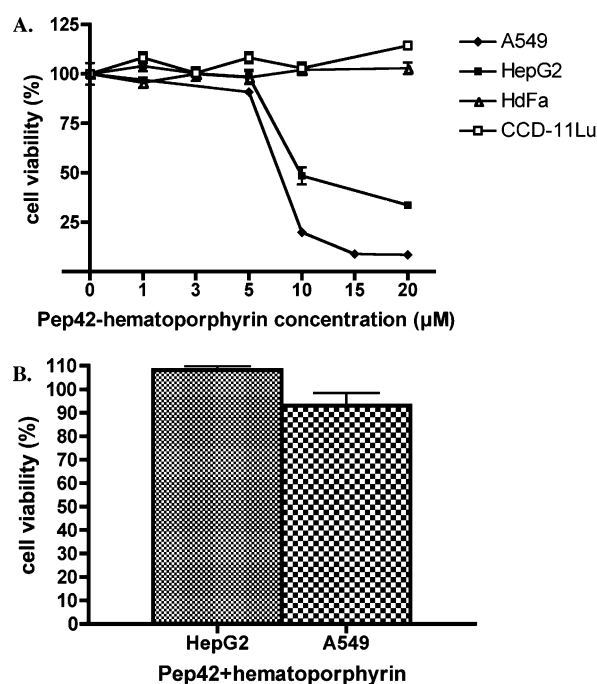


Figure 7. Phototoxicity of Pep42–hematoporphyrin conjugates toward surface expressing GRP78 cancer cells. The viability of the cells was assessed by a MTT assay. Results are expressed as percentages of cells still viable relative to nontreated control cells. (A) Phototoxicity of Pep42–hematoporphyrin conjugate on A549 cells and HepG2 cells. HdFa and CCD-11Lu cells served as controls. (B) An equimolar mixture of 10 μ M hematoporphyrin and Pep42 had no effect on HepG2 and A549 cells.

A549 cells, while HepG2 and A549 cells were unaffected by 10 μ M of an equimolar mixture of Pep42 and hematoporphyrin (Figure 7B).

The peptide without any cargo displayed maximum internalization levels after 0.5–1 h of incubation and the maximum colocalization with lysosomes occurred after 4 h; after 6 h the peptide was degraded by enzymes of the lysosomal compartment (Supporting Information, Figure S5). These observations clearly suggest that the peptide is stable enough to reach its target site in order to deliver a putative cargo. Furthermore, Pep42 can be cleared from lysosomes without toxic effects. At the same time, when Pep42 was conjugated to hematoporphyrin, the conjugate reached a maximum internalization 3–6 h after incubation. Similar results have been reported for other peptide–cargo conjugates.^{32,33} The observations indicate that the kinetics of cellular uptake are also dependent on the physicochemical character of the cargo.

Quantum Dot Conjugated Pep42 Site-Directed Transport in Xenograft Mice. Finally, in order to evaluate the

ability of Pep42 to find its target and carry a payload into the tumor tissue *in vivo*, Pep42 was conjugated to QDots and injected iv in the tail vein of tumor-bearing mice. Our results demonstrate that Pep42–QDots are able to home into the tumor tissue of living mammals (Figure 8A) while linear Pep42–QDots did not accumulate in the tumor (Figure 8D). The Pep42–QDots showed homing specificity for the relevant tumor tissue as accumulation of fluorescence within the targeted tissue was observed. Significantly less homing of cyclic Pep42–QDots was seen in relevant control organs, i.e., liver and spleen (Figure 8B and Figure 8C), and homing was not observed for linear Pep42–QDots (Figure 8E,F). These data provide evidence that Pep42 indeed has the ability to deliver cargo, in this case of significant size, specifically into the tumor tissue *in vivo* without accumulation in other organs, which is one prerequisite for any successful new approach in cancer chemotherapy.

Discussion

In this study, we report how a previously identified oligopeptide Pep42 binds specifically to overexpressed GRP78 on the surface of a number of human cancer cells. This specificity is then utilized to deliver cytotoxic drugs into malignant cells *in vitro* and also used as a tumor tissue homing beacon *in vivo*.¹⁶ We note that several linear chimeric peptides composed of GRP78 binding motifs have been identified to date, however, Pep42 is a cyclic 13-mer peptide that possesses a unique sequence, which does not share any known motifs with other GRP78-binding or otherwise tumor-homing peptides.¹⁷ Moreover, our screen was conducted using a phage display biopanning protocol on intact cells, thus ensuring that the selected ligands bound to the cellular receptor in its native context.³⁴

Small peptidic ligands are being considered as promising new tools for the delivery of cytotoxic drugs.³⁵ Although the mechanism of cellular uptake of canonical cell-penetrating peptides and other small peptidic ligands has not been fully understood in detail yet, it is believed that these types of peptides enter cells rapidly in a receptor-independent fashion.³⁶ In contrast our study provides strong evidence that receptor-mediated translocation is involved in the internalization of Pep42 and that clathrin-mediated endocytosis is the main mechanism of uptake. This would be in accordance with recent data that assume more than one mechanism for peptide-mediated intracellular delivery of various types of molecules and particulate structures.³

Pep42–QDots in combination with inhibitors of endocytosis and specific organelle markers for live cells were used for the investigation of the mechanism of endocytosis and colocalization experiments. The uptake mechanism of Pep42 into cancer cells involves clathrin-coated vesicles, which then

(32) Silhol, M.; Tyagi, M.; Giacca, M.; Lebleu, B.; Vives, E. *Eur. J. Biochem.* **2002**, 269 (2), 494–501.

(33) De Coupade, C.; Fittipaldi, A.; Chagnas, V.; Michel, M.; Carlier, S.; Tasciotti, E.; Darmon, A.; Ravel, D.; Kearsey, J.; Giacca, M.; Cailler, F. *Biochem. J.* **2005**, 390 (Part 2), 407–18.

(34) Oyama, T.; Rombel, I. T.; Samli, K. N.; Zhou, X.; Brown, K. C. *Biosens. Bioelectron.* **2006**, 21 (10), 1867–75.

(35) O'Connor, K. A.; Roth, B. L. *Nat. Rev. Drug Discovery* **2005**, 4 (12), 1005–14.

(36) Rhee, M.; Davis, P. *J. Biol. Chem.* **2006**, 281 (2), 1233–40.

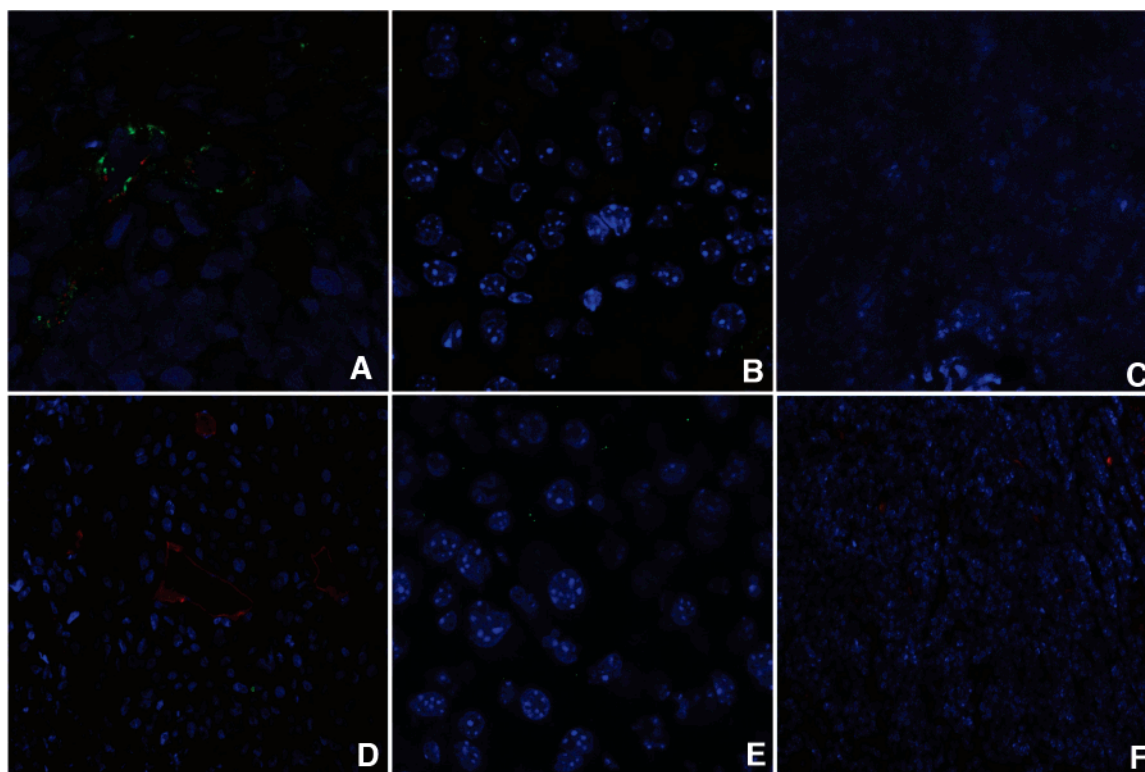


Figure 8. *In vivo* targeting of Pep42–QDot conjugates. Peptide-coated QDots were injected into the tail vein of tumor-bearing CB17Scid male mice, and the presence of QDots in the tumor and control tissue was assessed by examining sections under a confocal microscope. Accumulation of conjugate was observed in the tumor tissue with cyclic peptide QDots (A), and less material was found in the liver (B) and the spleen (C). Linear peptide coated QDots did not accumulate in the tumor (D), and also not in the control organs liver (E) and spleen (F). Blood vessels were stained with anti-CD31 (red).

transport the internalized peptide into the lysosomal compartment of the cell. Interestingly, the colocalization data presented here amends our previous findings suggesting that Pep42 is able to colocalize with the endoplasmic reticulum. Although the exact mechanism of endocytosis and subsequent intracellular fate may in fact depend on the particular cell type and cargo, the study presented here reveals two key steps of Pep42's cellular internalization, namely, an absolute required binding to the surface GRP78 and uptake via endocytosis.¹⁶ Importantly, artifacts of cellular peptide internalization induced by fixation, which are known in the literature,^{37,38} were avoided by only using live cells for the uptake studies of the fluorescent Pep42 into cells via confocal microscopy.

To evaluate the drug delivery of Pep42, we chose drugs with vastly different biochemical properties as well as different mechanisms of action. One drug was the amphipathic peptide D-(KLAKLAK)₂, an apoptosis-inducing drug, whose primary action site is the mitochondrial membrane, while the other drug consisted of hematoporphyrin, a photosensitizer that is active in the lysosomal compartment. Our data illustrate that the two drugs are not imported into any cell line tested without the appropriate drug delivery

system, namely, Pep42. Furthermore, conjugation of D-(KLAKLAK)₂ to Pep42 generated a reagent that selectively killed Me6652/4 and SJSA-1 in a surface level GRP78-dependent manner.

Clear evidence is presented of apoptosis induction in the cells treated with the conjugate, which strongly indicates that Pep42 allowed for the intracellular delivery and subsequent mitochondrial membrane targeting of D-(KLAKLAK)₂. The resistance observed of A549 and HepG2 cells to the D-(KLAKLAK)₂-induced killing might be the result of a cell type-dependent inability of the internalized peptide to access the cytoplasm and/or activate cell death-triggering mechanisms. Nevertheless, these cell lines were found to be susceptible to the Pep42–hematoporphyrin conjugate. To harness its full phototoxic activity, a localization into the lysosomal compartment of cancer cells is required.³¹ Applying the data gathered from the colocalization experiments of Pep42, this strategy resulted in successful killing of the two tumor cell lines, while no adverse effects were observed for the control cells.

(37) Lundberg, M.; Johansson, M. *Biochem. Biophys. Res. Commun.* **2002**, 291 (2), 367–71.

(38) Richard, J. P.; Melikov, K.; Vives, E.; Ramos, C.; Verbeure, B.; Gait, M. J.; Chernomordik, L. V.; Lebleu, B. *J. Biol. Chem.* **2003**, 278 (1), 585–90.

Nanoparticles are thought to have potential as novel intracellular probes for both imaging and drug delivery.^{39,40} Our results suggest that Pep42–QDots possess homing specificity for the relevant tumor tissue and significantly less homing in the control organs. Although the nanoparticles used in this study are rather simple, the use of more sophisticated systems could be envisioned for the future. Such devices may, as an example, deliver a drug to the site of action, release the drug, and report back its delivery. These findings highlight that Pep42 may serve as a promising candidate for future anti-cancer drug delivery platforms.

In conclusion, Pep42 can be considered as a promising tumor-targeting drug delivery system as it possesses the ability to deliver drugs with vastly different mechanisms of action into human cancer cells. We have shown that GRP78 is the exclusive receptor for Pep42, its mechanism for internalization and site directed transport in xenograft mice. We believe the data obtained warrant future evaluation of Pep42's drug delivery capabilities *in vivo* which will likely be more representative of clinically relevant aspects for the treatment of disease in human cancer.¹⁷ Overexpressed GRP78 on cancer cells can be selectively targeted by a cell internalizing peptide, and studies demonstrating the selective delivery of payloads for the killing of tumor cells will be the subject of future investigations.

Abbreviations Used

siRNAs, small interfering ribonucleic acids; GRP78, glucose-regulated protein 78; HSP 70, heat shock protein

- (39) Steiniger, S. C.; Kreuter, J.; Khalansky, A. S.; Skidan, I. N.; Bobruskin, A. I.; Smirnova, Z. S.; Severin, S. E.; Uhl, R.; Kock, M.; Geiger, K. D.; Gelperina, S. E. *Int. J. Cancer* **2004**, *109* (5), 759–67.
- (40) Kelly, K. A.; Nahrendorf, M.; Yu, A. M.; Reynolds, F.; Weissleder, R. *Mol. Imaging Biol.* **2006**, *8* (4), 201–7.

70; QDots, quantum dots; DMEM, Dulbecco's modified Eagle's medium; Me6652/4, human melanoma cell line; A549, human lung adenocarcinoma cell line; SJSA-1, human osteosarcoma cell line; HepG2, human hepatoma cell line; CCD-11Lu, human lung fibroblast cell line; EMEM, Eagle's modified minimum essential medium; HDFa, human dermal fibroblasts; FBS, fetal bovine serum; FITC, fluorescein isothiocyanate; EDCI, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide; DMSO, dimethyl sulfoxide; EDTA, ethylene diamine tetraacetate; SDS–PAGE, sodium dodecyl sulfate polyacrylamide gel electrophoresis; HRP, horseradish peroxidase; BSA, bovine serum albumin; PBS, phosphate-buffered saline; MCD, methyl- β -cyclodextrin; pAb, polyclonal antibody; CTB, cholera toxin subunit B; Ahx, aminohexanoic acid; Bpa, benzoylphenylalanine; SOBAG, super optimal broth ampicillin glucose; Fmoc, 9-fluorenylmethyl carbamate; dPEG6, discrete polyethylene glycol 6.

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Supporting Information Available: Figures S1–S5 depicting a schematic representation of analogues and conjugates of Pep42, identification of GRP78 on the cell surface, cell viability as a function of time and of Pep42–D-(KLAKLAK)₂ concentration, and colocalization of Pep42–QDots 525 endosomes with lysosomes in living cells. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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